

# Inhibition of glycine currents by dextromethorphan in neurones dissociated from the guinea-pig nucleus tractus solitarii

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- 1 The effect of dextromethorphan (DM) on the current induced by glycine in acutely dissociated nucleus tractus solitarii (NTS) neurones of guinea-pigs was studied by use of the whole-cell patch clamp technique. The effect of DM on  $\gamma$ -aminobutyric acid (GABA)-induced currents ( $I_{GABA}$ ) was also examined.
- **2** DM inhibited 30  $\mu$ M glycine-induced current ( $I_{Gly}$ ), without affecting the current caused by 30  $\mu$ M GABA. The action of DM was concentration-dependent, with a maximum effect at 100  $\mu$ M, and reversible. The half-maximum inhibitory concentration (IC<sub>50</sub>) of DM was 3.3  $\mu$ M, about 85 times higher than that of strychnine.
- 3 DM 3  $\mu$ M shifted the concentration-response curve for glycine to the right without affecting the maximum value. DM 10  $\mu$ M shifted the curve even more to the right, although it was not a parallel shift. Strychnine at a concentration of 0.1  $\mu$ M shifted the curve for glycine in a nearly parallel fashion.
- 4 The effect of 10  $\mu$ M DM was slightly weak voltage-dependency, but the lower concentration of DM, 3  $\mu$ M, inhibited  $I_{Gly}$  equally at -50 mV and +50 mV. The effect of 3  $\mu$ M DM on  $I_{Gly}$  showed no use-dependence. Blockade by strychnine 0.1  $\mu$ M showed no voltage- or use-dependence.
- 5 The results indicate that DM inhibits  $I_{Gly}$  in single neurones of NTS, and further suggest that DM at a low concentration may act on the glycine receptor-ionophore complex, but not on the Cl<sup>-</sup> channel of the complex. However, a relatively high concentration of DM may at least partly affect the Cl<sup>-</sup> channel of the complex.

**Keywords:** Dextromethorphan; glycine; nucleus tractus solitarii; whole-cell recording; dissociated neurones; guinea-pig; antitussive

#### Introduction

Glycine and  $\gamma$ -aminobutyric acid (GABA) are generally considered to be inhibitory neurotransmitters in the mammalian central nervous system (CNS). Although findings about the pharmacological significance of GABA receptor-ionophore complexes in the brain have accumulated (Schult & MacDonald, 1981; Olsen et al., 1982; Ticku & Rastogi, 1986; Yakushiji et al., 1989), much remains unknown about the pharmacological and pathophysiological significance of the glycinergic transmission in the CNS. We found that the intensity of cough responses was strengthened in guinea-pigs, when a minute amount of glycine was injected directly into the nucleus tractus solitarii (NTS) and the adjacent region where the cough reflex pathway is located (Honda et al., 1990). Furthermore, toxic doses of antitussives often facilitate convulsions in experimental animals (Takahama et al., 1978), possibly due to blockade of the receptors of inhibitory neurotransmitters such as glycine and GABA. Therefore, it seems of interest to determine whether antitussives influence the glycine-mediated response in the CNS.

In this study, we investigated the effect of an antitussive, dextromethorphan (DM), on  $I_{\rm Gly}$  in single neurones acutely dissociated from the NTS of guinea-pigs; strychnine was used as a reference drug. Since it has also been shown that GABA-ergic mechanisms participate in the effect of antitussives acting on the cough centre (Bolser *et al.*, 1994), the effect of DM on GABA-induced current ( $I_{\rm GABA}$ ) in the NTS neurones was also examined.

# Preparation

Single neurones of NTS were acutely dissociated from 7- to 10-day-old Hartley guinea-pigs according to the method previously described (Huguenard & Alger, 1986). The guinea-pigs were decapitated under ether anaesthesia. The brain was quickly removed and dissected into slices 400  $\mu m$  in thickness by a microslicer (DTK-1000, D.S.K.). The slices were pre-incubated in external solution bubbled with 100%  $O_2$  at 37°C for 30 min. Thereafter, the NTS region was dissected out by punching the slices with a small hand-held needle. The small tissue chunks including the NTS were stirred in external solution containing 1.8 mg ml $^{-1}$  trypsin for 30–60 min at 37°C with constant oxygenation, then triturated with a fire-polished micro-pipette.

The ionic composition of external solution was (in mM): NaCl 140, KCl 5, CaCl<sub>2</sub> 1, MgCl<sub>2</sub> 1, N-2-hydroxyethyl-piperazine-N'-2-ethanesulphonic acid (HEPES) 10 and D-glucose 24. The pH was adjusted to 7.4 with tris(hydroxymethyl) aminomethane (Tris-OH). The patch-pipette solution contained (in mM): KCl 140, CaCl<sub>2</sub> 1, ATP-Mg 2, ethylene glycolbis (β-aminoethyl ether)-,N,N,N',N'-tetraacetic acid (EGTA) 10 and HEPES 10. The pH was adjusted to 7.2 with Tris-OH. In the current-voltage relationship experiment, K<sup>+</sup> in the pipette solution was replaced with equimolar Cs<sup>+</sup>, to suppress the voltage-dependent K<sup>+</sup>-currents.

## Electrical measurements

Electrical measurements were made with the whole-cell configuration mode of the patch clamp technique (Hamill *et al.*, 1981). The resistance between the recording electrode filled with the patch-pipette solution and the reference electrode in the external solution was  $4-6~\mathrm{M}\Omega$ . Currents were measured

Methods

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with a patch-clamp amplifier (Axopatch 1D, Axon Instrument), and monitored on both an oscilloscope (MS-5021, Iwatsu) and a pen recorder (Model 3021, Yokogawa). Data were filtered at 1 kHz, digitized, and stored on microcomputer disk for subsequent analysis by use of the pClamp system (Axon Instruments).

#### Drugs

Drugs dissolved in the external solution were applied by the 'Y-tube' technique that allowed complete exchange of the solution within 20 ms as described by Murase *et al.* (1989). The drugs and chemicals used in the present experiments were from commercial sources.

#### Statistical analysis

Experimental values are given as  $mean \pm s.e.mean$ . Unpaired Student's t test was used to assess the significance of difference when necessary, and a P value of less than 0.05 was considered to be significant.

### Results

### Effect on glycine- and GABA-induced current

In freshly dissociated NTS neurones, which were voltage-clamped to a holding potential ( $V_{\rm H}$ ) of -50 mV, glycine induced an inward current in a concentration-dependent manner at concentrations of 3  $\mu{\rm M}$  to 3000  $\mu{\rm M}$ . The glycine-induced current ( $I_{\rm Gly}$ ) gradually desensitized during a continuous application of glycine lasting 20 s. Glycine was applied every 1 min, at which interval no desensitization of the peak current occurred. The  $I_{\rm Gly}$  induced by glycine 30  $\mu{\rm M}$  in the NTS neurones of guinea-pigs was completely depressed by strychnine 1  $\mu{\rm M}$  but not by bicuculline 10  $\mu{\rm M}$  at all (data not shown).

DM at a concentration of 100  $\mu$ M produced no response in the NTS neurones. But it depressed the current induced by glycine 30  $\mu$ M (Figure 1a). The depressive effect was concentration-dependent over the range 0.1  $\mu$ M to 100  $\mu$ M (Figure 1b), with complete depression at 100  $\mu$ M. The half-maximum inhibitory concentration (IC<sub>50</sub>) of DM on the current induced by 30  $\mu$ M glycine was 3.3  $\mu$ M. On the other hand, strychnine strongly blocked  $I_{\rm Gly}$  at 0.001  $\mu$ M to 1  $\mu$ M, with an IC<sub>50</sub> of 0.039  $\mu$ M, about 85 times lower than that for DM. DM at 100  $\mu$ M had no effect on the current induced by GABA 30  $\mu$ M ( $I_{\rm GABA}$ ) (Figure 1a) in 5 neurones examined, while  $I_{\rm GABA}$  was depressed by bicuculline 10  $\mu$ M (data not shown).

#### Effect on concentration-response curve for glycine

To study the mechanism of the inhibitory action of DM on  $I_{\rm Giy}$ , the effect of DM on the concentration-response relationship for glycine was examined. As shown in Figure 2, DM 3  $\mu$ M slightly shifted the concentration-response curve for glycine to the right without depressing the maximal response of glycine. DM 10  $\mu$ M shifted the curve even more to the right, although it appeared not to be a parallel shift. The EC<sub>50</sub> values in the presence and absence of DM 3  $\mu$ M were 67  $\mu$ M and 39  $\mu$ M, respectively, and the value in the presence of DM 10  $\mu$ M was 120  $\mu$ M. Strychnine also shifted the curve to the right in a nearly parallel fashion. The EC<sub>50</sub> in the presence of 0.1  $\mu$ M strychnine was 200  $\mu$ M.

#### Effect on current-voltage relationship for glycine

Figure 3 shows the current-voltage (I–V) relationship for glycine in the presence and absence of DM 10  $\mu$ M. In the absence of DM,  $I_{\rm Gly}$  reversed direction at +10 mV, and the I-V relationship was almost linear. The reversal potential for  $I_{\rm Gly}$  was close to the Cl<sup>-</sup> equilibrium potential (+1.2 mV) calculated by Nernst equation from the given intra- and extracellular

Cl<sup>-</sup> concentrations and their Cl<sup>-</sup> activities, indicating that the currents induced by glycine were carried by Cl<sup>-</sup>. DM depressed the  $I_{\rm Gly}$  at different V<sub>H</sub>. In the presence of DM 10  $\mu$ M, the I-V relationship for glycine did not show complete linearity; the  $I_{\rm Gly}$  at a V<sub>H</sub> of -50 mV was reduced by  $71.4\pm2.8\%$  while that at a V<sub>H</sub> of +50 mV by  $46.4\pm3.3\%$ . There was a statistically significant difference between the two values. This result suggests that the inhibitory action of DM 10  $\mu$ M on  $I_{\rm Gly}$  is weakly

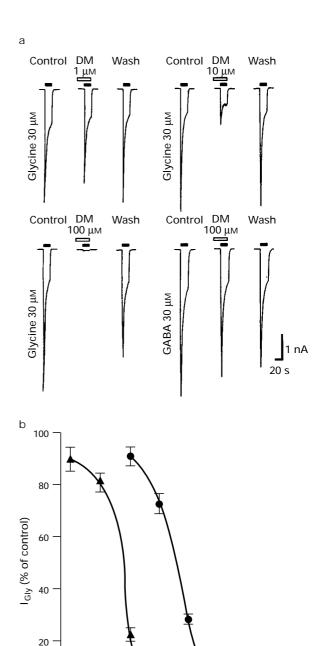


Figure 1 Concentration-dependent inhibitory effect of dextromethorphan (DM) and strychnine in NTS neurones. (a) A neurone was voltage-clamped at -50 mV. DM depressed 30 μM glycine-induced Cl<sup>−</sup> current  $I_{\rm Gly}$  in a concentration-dependent manner, but did not change the current induced by 30 μM GABA. An apparent depression of the  $I_{\rm GABA}$  was due to a run-down of the response. All responses were obtained from the same neurone. (b) Concentration response curve for the inhibitory effects of DM (●) and strychnine (▲) on  $I_{\rm Gly}$ . Each point denotes mean from 4−5 experiments; vertical lines show s.e.mean.

1

10

100

0.01

0.001

0.1

Antagonist concentration (µм)

voltage-dependent. However, in the presence of DM 3  $\mu$ M, the  $I_{\rm Gly}$ s at -50 mV and +50 mV  $\rm V_H$  were reduced to the same extent (data not shown), and the line through the two values crossed the abscissa scale at +3.6 mV (data not shown), close to the calculated  $\rm Cl^-$  equilibrium potential. This result suggests that the action of DM at a low concentration might be not voltage-dependent. On the other hand, strychnine at  $0.1~\mu$ M depressed  $I_{\rm Gly}$  by about 65% at different  $\rm V_{HS}$  of -70 to +50 mV, i.e., no distortion of the I-V relationship for glycine was observed. The result indicates that the inhibitory action of strychnine on  $I_{\rm Gly}$  is not voltage-dependent.

Effect of DM on response to repeated application of glycine

Further experiments were undertaken to see whether the effect of DM is use-dependent. As shown in Figure 4, the magnitude

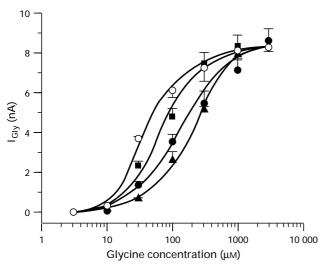
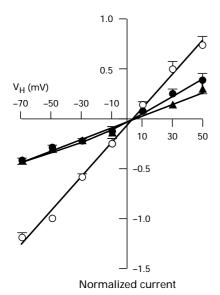


Figure 2 Concentration-response curves for the inhibitory effect of glycine on  $I_{\rm Gly}$  in the absence and presence of dextromethorphan (DM) or strychnine. Neurones were voltage-clamped at -50 mV. Each point denotes mean and vertical lines s.e.mean from 4-9 experiments. ( $\bigcirc$ ) Effect of glycine alone, ( $\blacksquare$ ) with DM 3  $\mu$ M, ( $\bullet$ ) with DM 10  $\mu$ M, ( $\blacktriangle$ ) with strychnine 0.1  $\mu$ M.

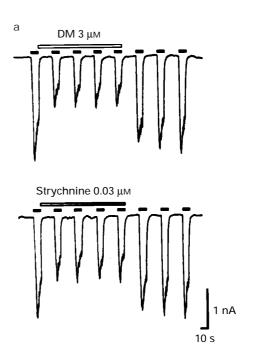


**Figure 3** Current-voltage (*I*-V) relationship for  $I_{\rm Gly}$  in the absence or presence of dextromethorphan (DM) 10  $\mu$ M or strychnine 0.1  $\mu$ M. All responses were normalized to the peak current induced by glycine 30  $\mu$ M at a V<sub>H</sub> of -50 mV. Each point denotes mean and vertical lines s.e.mean from 4-14 experiments. ( $\bigcirc$ ) Effect of glycine alone, ( $\bigcirc$ ) with DM 10  $\mu$ M, ( $\triangle$ ) with strychnine 0.1  $\mu$ M.

of the inhibitory action of DM 3  $\mu$ M was not changed by repeated application of glycine in the continued presence of DM. A similar result was obtained in the presence of a low concentration of strychnine.

#### Discussion

This manuscript provides the first evidence that DM potently inhibits  $I_{\rm Gly}$  in brain neurones. DM at a concentration of 100  $\mu{\rm M}$  did not have an effect on the current caused by 30  $\mu{\rm M}$  GABA (GABA concentration near the  $K_{\rm D}$  value for NTS neurones of guinea-pig, data not shown).



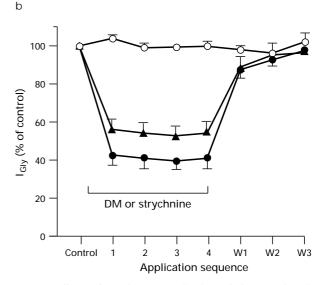


Figure 4 Effects of continuous application of dextromethorphan (DM) and strychnine on  $I_{\rm Gly}$ s induced by 4 successive applications of glycine. (a) Typical current record of  $I_{\rm Gly}$ s in the continuous presence of DM 3  $\mu$ M or strychnine 0.03  $\mu$ M. Glycine 30  $\mu$ M was applied for 10 s every 30 s. V<sub>H</sub> was -50 mV. (b) Summarized data of the effects of continuous presence of DM or strychnine on  $I_{\rm Gly}$ s. Concentrations of DM and strychnine, applied for the period indicated by a bar in the figure, were 3  $\mu$ M and 0.03  $\mu$ M, respectively. Each point denotes mean and vertical lines s.e.mean (n=4-6) of percentages of control response. Effects of ( $\bullet$ ) DM, ( $\blacktriangle$ ) strychnine and ( $\bigcirc$ ) vehicle are shown. W1–W3: washing.

DM in the range  $10-100~\mu\text{M}$  inhibits voltage-dependent  $\text{Ca}^{2+}$  and  $\text{Na}^+$  channels (Tortella *et al.*, 1989; Lodge & Johnson, 1990; Netzer *et al.*, 1993; Trube & Netzer, 1994). However, it is unlikely that the effect of DM on  $I_{\text{Gly}}$  is produced through an action on these channels.  $I_{\text{Gly}}$  is carried by  $\text{Cl}^-$  in various neurones of the CNS (Bormann *et al.*, 1987; Ito *et al.*, 1991; Uneyama *et al.*, 1993); including the NTS neurones of guinea-pigs.

[ ${}^{3}$ H]-DM has at least two distinct binding sites in guinea-pig brains, a high-affinity site ( $K_{\rm D} = 13 - 20$  nM) and a low-affinity site ( $K_{\rm D} > 200$  nM). High affinity [ ${}^{3}$ H]-DM binding was found not to be displaced by various neurotransmitters including glycine and GABA (Craviso & Musacchio, 1983). Furthermore, because the  $K_{\rm D}$  for the high affinity site is in the nM range, far smaller than the the concentration needed to block  $I_{\rm Gly}$ , it is unlikely that DM inhibits  $I_{\rm Gly}$  through an action on the high affinity site.

Of the two glycine-binding sites in the CNS, one is strychnine insensitive and part of the N-methyl-D-aspartate (NMDA) receptor (Johnson & Ascher, 1987; Harris & Miller, 1989; Kessler et al., 1989; Yamada et al., 1989; Kemp & Leeson, 1993). Netzer et al. (1993) have shown that DM depresses the NMDA-induced current in cultured cortical neurones of the rat. The depression was not due to competitive blockade of the strychnine insensitive glycine binding site but due to blockade of the ion channel activated by NMDA and glycine (Netzer et al., 1993). Therefore, DM does not interfere with the strychnine insensitive glycine binding site in a competitive fashion.

There are a few studies that suggest a non-competitive mode of action for strychnine blockade of glycine responses in the brain (Krishtal *et al.*, 1988; Akaike & Kaneda, 1989) and in *Xenopus* oocytes injected with rat brain mRNA (Houamed *et al.*, 1984). However, many findings have shown that the strychnine antagonism is competitive, especially in neurones of the lower brain stem and spinal cord. In the present study, strychnine 0.1  $\mu$ M shifted the concentration-response curve for glycine to the right in a nearly parallel fashion without affecting the maximum value. This result suggests that the action of strychnine in NTS neurones of guinea-pigs is probably competitive. DM also shifted the curve for glycine to the right. However, the rightwards shift appeared not to be in a typical

parallel fashion, although the maximum  $I_{\rm Gly}$  response was not affected. Thus, it seems to be difficult to conclude simply that DM inhibits  $I_{\rm Gly}$  in a competitive manner, especially with regard to the action of 10  $\mu$ M DM.

The block of the NMDA receptor by DM is thought to be voltage- and use-dependent, although direct evidence for this has not been published. With respect to this, the block of the NMDA receptor by dextrorphan, a closely related morphinan, is voltage- and use-dependent (Parsons *et al.*, 1993). Therefore, we examined the voltage- and use-dependency of the inhibitory action of DM on  $I_{\rm Gly}$ . Interestingly, the effect of a low concentration of DM on  $I_{\rm Gly}$  was neither voltage-dependent nor use-dependent, although the action of DM 10  $\mu$ M showed a very weak voltage-dependency. The results suggest that DM in a low concentration may act on the glycine receptor-ionophore complex, but not on the C1<sup>-</sup> channel of the complex. However, a relatively high concentration of DM may at least partly affect the Cl<sup>-</sup> channel of the complex.

It seems possible that the inhibitory action on  $I_{\rm Gly}$  might be associated with antitussive drugs, because our recent work revealed that various antitussives including narcotic antitussives such as codeine inhibit  $I_{\rm Gly}$  in NTS neurones without depressing  $I_{\rm GABA}$  (Otsuka *et al.*, 1996). However, further studies are needed to clarify whether it is the antitussive or the adverse effects of these drugs that is linked to the action on glycine receptors. Bolser *et al.* (1994) suggested that an involvement of GABA<sub>B</sub> receptors in the mechanism of action for centrally-acting antitussive drugs.

Finally, the present results may partly explain the mechanism of the ameliorating action of DM on the neurological symptoms of nonketotic hyperglycinemia (Schmitt *et al.*, 1993). In this condition, the glycine concentration in plasma and cerebrospinal fluid is elevated (Nyhan, 1989) and the pathophysiological effects of this disease are attributed to the inhibitory action of glycine at the postsynaptic, strychninesensitive receptor (Gitzelmann *et al.*, 1977).

We sincerely thank Drs B.E. Alger and T. Shirasaki for their kind guidance in a dissociation technique of brain neurones and a 'Y-tube' technique, respectively. We are also grateful to Professor Work for reading the manuscript.

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(Received August 21, 1996 Revised October 18, 1996 Accepted October 31, 1996)